The amount of each solution was equal to the amount of water containing the colony. Each solution was added drop by drop very slowly. Gradually some of the solution containing the colony was removed in order to keep the amount constant.

The time required for the application of each solution of chloretone varied from 15 to 30 minutes. After the colony had been in the saturated chloretone solution for 15 minutes, the killing agent was added.

A 3% solution of formalin was diluted with a saturated solution of chloretone, and the following grades were used:

1. 1 part 3% sol. of formalin to 2 parts sat. sol. of chloretone.
2. 1 part 3% sol. of formalin to 1 part sat. sol. of chloretone.
3. 2 parts 3% sol. of formalin to 1 part sat. sol. of chloretone.
4. 3% formalin.

These solutions were added drop by drop in the same manner as for narcotization, and 15 to 30 minutes were allowed for the application of each grade.

Two and one half to five hours are necessary for the entire procedure. For cristatella the minimum time is sufficient, but for Plumatella and especially for Fredericella the maximum time is necessary.

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CELLULOID COVERS FOR LARGE MICROSCOPIC SLIDES

This method is devised to meet the need of covering serial sections of large objects, such as advanced embryos. One can reach, for examination with a compound microscope, every point on slides measuring 7x3 inches. These may be cut from double thickness window glass.

Such material as will not permit of being stained in bulk before embedding can be stained on the slide in a photographic tray of hard rubber (not painted). Glass covers are not practicable on such slides; but thin window glass may be used where high powers are not to be employed. We have recently found that high powers, including oil immersion lenses, may be used on these large slides by covering with what is known among the dealers in photographic sup-

plies as *tissue celluloid* sheets. These come in sizes to cover three of the slides of dimensions given above at a cost of fifteen or twenty cents for three.

These celluloid covers do not seem to show any dissolving action on the part of the xylol in the balsam, although quite thin balsam was used on the slides, some of which have been mounted two months or longer. The celluloid sometimes falls into gentle undulations but not of such a nature as to distort the image seen with high powers. The celluloid does have a tendency to squeeze the balsam out at the edges where it curls up. This would make it impractical for substitution for the ordinary sizes of covers although it might be found much cheaper. Whether the celluloid will become darkened after a time as do the sheets of it used in shades and curtains of automobiles remains to be seen. Perhaps it would be well to keep it in the dark.

CHAS. BROOKOVER.

MICA SHEETS FOR CARRYING HISTOLOGICAL SECTIONS

Mica sheets may be used with Mayer's egg albumen adhesive for carrying sections of paraffin material through the staining process, after which the mica can be cut into pieces and issued to students from xylol or creosote ready for mounting in balsam.

This method has been used in Germany, but has not been employed to the extent it deserves in this country. When the mica sheets procurable at the stove dealers or hardware store is split into thin flakes and the fresh surface used, no difficulty is experienced in getting the sections to adhere. For warming and floating the sections out smooth, the mica was cut into sizes to fit over a slide. The remainder of the process of staining and clearing was done with the mica sheet alone. For a moderate number of students in a class, a specimen for each can be stained as quickly as a single slide can be handled. It is well to mount in the balsam with the specimen uppermost as there may be air spaces between the laminæ of the mica.

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